DOI: 10.22620/agrisci.2024.41.004

SILICON APPLICATION UNVEILED: A REVIEW OF INSIGHTS INTO PLANT DEFENSE MECHANISMS UNDER BIOTIC CHALLENGES

Adelina S. Harizanova

Agricultural University – Plovdiv, Bulgaria *Corresponding author's Email: a_harizanova@au-plovdiv.bg

Abstract

Silicon (Si) is the second most abundant element after oxygen in soil. Nevertheless, it was not considered essential for plant growth and development. In the last decades, many researchers have reported that silicon can mitigate to some extent the adverse effects of variable biotic and abiotic stresses caused by salinity, chilling, heating, nutritional imbalance, heavy metals, diseases, herbivores, and many others. There are different hypotheses regarding the mechanisms of the silicon mode of action. Some of them propose that the silicon treatment is probably related to the structural modification of the plant cells and tissues. Others speculate that silicon could be involved in plant metabolism. The interaction between the two mechanisms is also a very reliable hypothesis. Most of the studies focused on the influence of silicon on alleviating the negative effects of abiotic stress factors. The reports about the effect of silicon under biotic stress are not so abundant. Today, there is no doubt about the positive effect of silicon application in alleviating stress and reducing the pest and disease incidence and severity. However, the mechanisms of the silicon-induced plant responses are not yet completely clear. This motivated the current study to analyze the data presented about the silicon-induced alleviation of biotic stress.

Keywords: silicon, stress, disease, pest, physical barrier, plant metabolism

INTRODUCTION

Silicon (Si) is the second most abundant element after oxygen in the Earth's crust (Ma and Yamaji, 2006), yet it was not considered essential for plants (Arnon & Stout, 1939). In 2015, according to the International Plant Nutrition Institute (IPNI), Si was categorized as a "beneficial substance". The application of silicon-containing products can have several agronomic benefits, including improved plant growth and increased yield (Epstein, 1994, Richmond & Sussman, 2003) and protection from pests and diseases (Massey et al., 2006, Sun et al., 2022). Silicon is absorbed by the plant roots as soluble silicic acid Si(OH)₄ and in ionized form - Si(OH)₃O⁻, its which predominates at high pH of the soil solution (Currie & Perry, 2007). It is deposited in various plant organs and tissues as solid amorphous silica (SiO₂.nH₂O), where it interacts with polyphenols and pectin and enhances cell wall strength and rigidity (Jinger et al., 2017). These two compounds are found in soil at concentrations of 0.1 to 0.6 mM and are readily absorbed by plants (Epstein, 1994). Silicon accumulation varies between species, ranging from 0 to about 10% of the plant dry shoot weight, and its deposition is not the same in the different parts of the plant (Shwethakumari & Prakashand, 2018). The most significant amounts of Si are accumulated in the plants of the Poaceae, Equisetaceae and Cyperaceae families (Si > 4%) followed by the species belonging to the Cucurbitacea, Urticaceae and *Camelinaceae* families which accumulate medium amounts of Si (2% < Si < 4%) (Currie & Perry, 2007). Most of the other plant species

accumulate low quantities of Si. Depending on the ability to absorb Si, plants are divided into three groups: active accumulators, passive accumulators, and excluders (Guntzer et al., Hernandez-Apaolaza, 2012. 2014). McNaughton et al. (1985) suggested that silicon may be crucial in controlling pests. Until nowadays, Si has been shown to improve plant resistance and decrease plant damage caused by pathogens, insects, and non-insect pests. In the last decades, many researchers have proposed that silicon can, to some extent, mitigate the adverse effects of variable biotic and abiotic stresses. A significant number of studies were carried out with monocot plants, including rice (Yang et al., 2018, Roy et al., 2023, Tenguri et al., 2023), wheat (Singh et al., 2022, Ashfaq et al., 2023), barley (Sakr, 2021) and maize (Nascimento et al., 2018, Acevedo et al., 2021, Haq et al., 2022). Other articles investigate the Si effect in dicot plants belonging to various families, namely Cucurbitaceae (Savvas et al., 2009), Solanaceae (Liu et al., 2009, Dos Santos et al., 2015, Somapala et al., 2016), *Fabaceae* (Shwethakumari & Prakashand, 2018) and many others. Almost all of the studies declared reduced pest and disease severity. Still, not all of them explain the mechanism by which silicon reduces the adverse effects of stress.

Forms of silicon applied for biotic stress alleviation

Silicon is absorbed from the soil by the roots, but it can also be applied to the shoots via foliar spray. The forms of available silicon used in the field or in glasshouses are various (Table 1), including potassium silicate K₂SiO₃ (Schuerger & Hammer, 2003, 2006, 2007, Liu et al., 2020), sodium silicate Na₂SiO₃ (Li et al., 2009, Sun et al. 2022), calcium silicate CaSiO₃ (Resende et al., 2013, Alves et al., 2015), silicic acid H₄SiO₄ (Shwethakumari & Prakash, 2018), silica gel (SiO₂) (De Oliveira et al., 2020), and Si-rich straw or hulls (Somapala et al., 2016, Bakhat et al., 2020) (Table 1).

Form of Si	Plant	Reference	
Potassium silicate	strawberry	Abd-El-Kareem et al. (2019), Kanto et al.	
(K_2SiO_3)		(2007), Liu et al. (2020)	
	cucumber	Schuerger & Hammer (2003)	
	zinnia	Ranger et al. (2009)	
	lettuce	Garibaldi et al. (2012)	
Sodium silicate	wheat	Basagli et al. (2003)	
(Na_2SiO_3)	potato	Li et al. (2009)	
	strawberry	Abd-El-Kareem et al. (2019)	
	cucumber	Sun et al. (2022)	
Calcium silicate	sorghum	Resende et al. (2013)	
(CaSiO ₃)	sweet pepper	Alves et al. (2015)	
	rice	Linger et al. (2017)	
	strawberry	Abd-El-Kareem et al. (2019)	
Silicic acid (H ₄ SiO ₄)	soybean	Shwethakumari & Prakash (2018)	
Silica gel (SiO ₂)	vine	Parrilli et al. (2019)	
	wheat	De Oliveira et al. (2020)	
Silicon-rich biochar	tomato	Somapala et al. (2016)	
(rice husk)	wheat	Otitodun et al. (2017)	
	eggplant	Bakhat et al. (2020)	

Table 1. Forms of silicon applied for biotic stress alleviation

The beneficial effects of Si are associated with the mechanical and physiological modification of plants, depending on whether it is applied to the roots or to the shoots of *Podosphaera aphanis*-infested strawberry plants (Liu et al., 2020).

The protective role of Si application to the root system was attributed to Si accumulation in leaves, which hinders cuticle penetration by pathogens (Seal et al., 2018). According to several studies, the application of Si to the root system is promising in the protection against pathogens, but when supplied to the leaves, the protective effects are lower (Gomez et al., 2017).

According to other researchers, the mode of action of the foliar-applied K_2SiO_3 on powdery mildew could be attributed to the formation of physical barriers and osmotic effects on leaf surfaces (Rodrigues et al., 2009). Another cheap and natural source of Si is the Sirich plant material such as the rice hull (husk) (Otitodun et al., 2017). It is organic, rich in silicon and contains about 8% of Si in its dry weight (Somapala et al., 2016).

Stress responses in plants

During the process of evolution, plants have developed various specific mechanisms to overcome the adverse effects of different stressful factors (Rejeb et al., 2014). Exposure to biotic and abiotic stress causes a disruption in plant metabolism, which leads to physiological damage (Bolton, 2009) and a decrease in plant health and productivity. Abiotic stress is one of the most essential characteristics of growth and has a significant impact on it. Therefore, it is responsible for severe losses in the field. The resulting reduction in growth can reach > 50% in most plant species. Biotic stress is an additional obstacle inducing intense pressure on plants (Mordecai, 2011).

During plant-pathogen co-evolution, plants have developed a variety of defense mechanisms to prevent invasion and colonization by pathogens, belonging to diverse species including fungi, oomycetes, bacteria, viruses, and animals. A plant defense is successful when it ensures an early and fast restriction of the pathogen infestation and subsequent induction and mobilization of structural and biochemical protective mechanisms (Voigt, 2014).

Silicon-induced physical amendment of plant cells tissues

Plants can react to the attack directly and indirectly. Direct protections related to the morphological characteristics of the plant such as wax layer, trichomes (hairs), lignification of cell walls, affect the enemy feeding process (Dos Santos et al., 2015, Massey & Hartley, 2009). These plant features represent a physical (mechanical) barrier and are the first line of defense. Plant cell walls are the front line of defense against pests and pathogens (Swaminathan et al., 2022). One of the hypotheses about the silicon mode of action is related to the mechanical changes in the plant cell and tissues that increase their strength and abrasiveness. The physical defenses against herbivores consist of structures such as raphides (needle like crystals), trichomes (hairy structures), thorns, rough and tough epidermal cells, spines, hard shells and pods (Vicari & Bazely, 1993). Many studies report a decrease in the pathogen population due to a reduced penetration of pathogens in strawberry (Kanto et al. 2007, Liu et al. (2020), cucumber (Menzies et al., 1991), potato (Liu et al., 2009) and many others (Table 2 and Table 3).

The formation of a mechanical barrier in the cuticle and in the cell walls, caused by the polymerization of silicon was the first hypothesis about the Si mode of action in relation to reduction in plant disease severity (Samuels et al., 1994, Song et al., 2021). Silicon prevents the physical penetration of pathogens as it forms a thick layer under the leaf cuticle making the plant cell wall less susceptible to enzymatic degradation (Yoshida et al., 1962). Agricultural University – Plovdiv 🎇 AGRICULTURAL SCIENCES Volume 16 Issue 41 2024

Disease/pathogen	Plant host	Mode of action	Reference	
Fungi				
Powdery mildew	wheat	Synthesis of phenolics and phytoalexins	Rémus-Borel et al. (2005)	
	strawberry	Physical barrier	Kanto et al. (2007)	
	strawberry	Physical barrier, biochemical amendment	Liu et al. (2020)	
	cucumber	Physical barrier, papillae, phenolic synthesis	Menzies et al. (1991), Samuels et al. (1994)	
	cucumber	Flavonoid phytoalexins synthesis	Fawe et al. (1998)	
<i>Fusarium oxysporum</i> f. sp. cucumerinum	cucumber	Antioxidant defense, activation, photosynthesis increase, Calvin cycle related to the gene expression	Sun et al. (2022)	
Dry rot of potato tubers (<i>Fusarium sulphureum</i> Schltdl.)	potato	Thickening of the hyphal cell walls, cell distortion, and the deposition of electron- dense material in hyphal cells	Liu et al. (2009)	
Leaf blight (<i>Phomopsis</i> obscurans)	strawberry	Activation of peroxydase, polyphenol oxydase and chitinase	Abd-El-Kareem et al. (2019)	
Early blight (Alternaria solani)	Tomato	Expression of defense-related genes and antioxidant enzymes	Gulzar et al. (2021)	
Anthracnose	Tomato	Thicker cuticle and increased fruit firmness	Somapala et al. (2016)	

Table 2. Potential mechanisms of the silicon-induced protection against diseases (fungal diseases)

Table 3. Potential mechanisms of the silicon-induced protection against diseases (bacteria and viruses)

Disease/pathogen	Plant host	Mode of action	Reference	
Bacteria	·			
Bacterial blight		Activation of β -1,3-glucanase, endochitinase and exochitinase	Xue et al. (2010)	
(Xanthomonas oryzae pv. oryza)	rice	Increased synthesis of total soluble phenolics and lignin, activities of PAL and PPO	Song et al. (2016)	
Bacterial wilt (<i>Ralstonia solanacearum</i>)	sweet pepper	Increased production of chitinase, superoxide dismutase, ascorbate, peroxidase, β -1,3-glucanase, lignin and total protein	Alves et al. (2015)	
Bacterial wilt (<i>Ralstonia</i> solanacearum)	tomato	Priming effect, triggering the expression of JA, ET, and/or ROS dependent genes	Ghareeb et al. (2011)	
Viruses				
Tobacco ring spot virus (TRSV)	tobacco	Delay of TRSV systemic symptom formation	Zellner et al. (2011)	

Outside the cells, Si is accumulated in the subcuticular layer, the cell wall and intercellular spaces (Datnoff et al., 2007). The silicon-induced pathogen resistance is associated with a delayed incubation period, a reduced colony size, a decreased lesion size and number, and a suppressed inoculum production of fungi (Debona et al., 2017).

According to many authors, the plant resistance to pests could be enhanced by antixenosis (reducing pest colonization) and antibiosis (reducing the reproductive period and the fecundity of pests such as aphids) after Si fertilizer application (Ranger et al., 2009, Dias et al., 2014, Boer et al., 2019). In the research of de Oliveira et al. (2023), the soil application of silicon led to a linear increase in the productivity of wheat and to a reduction in Sitobion avenae number. Both wheat genotypes (susceptible and constitutive resistant) demonstrated enhanced productivity and decreased aphid numbers, indicating that the Si treatment is compatible with the use of resistant plants. The authors couldn`t explain whether the observed resistance was caused by antibiosis, antixenosis, or both, but the Si applications demonstrated the potential of using S. avenae in wheat in integrated management.

Silicon reduces the digestibility of tissues (Calandra et al., 2016, Massey et al., 2006). Silicification occurs in macro-hairs and typical rectangular epidermal cells (Hodson et al., 1984), which is common for many grasses. Many authors suggested that the reason for the silicon-induced reduction in the pest numbers was the physical amendment of the plant tissues (Acevedo et al., 2021, Dos Santos et al., 2015, Hag et al., 2022, Massey & Hartley, 2009, Roy et al., 2023, Sidhu et al., 2013, Tenguri et al., 2023, Vilela et al., 2014, White & White 2013, Xue et al., 2022, Yang et al. 2018). The silicon deposits were clearly located under the Scanning Electron Microscope at different doses of the silicon treatment, indicating that the increase in the silica dose enhanced its deposits, confirming its role in the defense mechanism (Roy et al., 2023).

Regarding the anatomy of their mouth parts, insect and noninsect pests are divided into piercing-sucking (like leafhoppers, aphids, whiteflies, thrips and mites) and chewing (like moths, butterflies, and beetles) (Kumar & Rathor, 2020). Both groups could be effectively suppressed after feeding on Si-treated plants (Table 4 and Table 5).

Insect Pest	Host plant	Mode of action	Reference	
Chewing pests				
Brown plant hopper (Nilaparvata lugens (Stål)	rice	Physical barrier	Yang et al. (2018), Roy et al., (2023), Tenguri et al. (2023)	
Rice leaf folder <i>Cnaphalocrocis medinalis</i> (Guenee)	rice	HIPVs production	Han et al. (2016)	
Stalk borer Diatraea saccharalis (F.)	sugarcane	Physical barrier	White & White (2013), (Sidhu et al., 2013), Vilela et al. (2014)	
Fall armyworm <i>Spodoptera</i> <i>frugiperda</i> (Smith)	maize	Physical barrier	Nascimento et al. (2018), Acevedo et al. (2021), Haq et al. (2022)	
African armyworm Spodoptera exempta (Walker)	ryegrass	Physical barrier	Massey & Hartley, (2009)	
Tomato leaf miner <i>Tuta</i> <i>absoluta</i> (Meyrick)	tomato	Physical barrier	Dos Santos et al. (2015)	

Table 4. Potential mechanisms of the silicon-induced protection against insect pests (chewing insects)

According to Massey & Hartley, (2009) the high content of silicon in tissues leads to a reduced leaf digestibility and causes mandible wear, which is probably crucial for the herbivore performance. The authors reported that the Si-rich diet was responsible for the extremely rapid insect mandibles wear, occurring within a single instar. The damage of the mandibles correlated with the decreased absorption of nitrogen and reduced growth rates. Herbivores, fed on Si-rich leaves, cannot completely recover after switching diets. This demonstrates the strong deterrent effect of the silica-based defenses. These defenses, in contrast to many chemical defenses, cannot be compensated or overcome with a dietary change: the adverse effects remain even when the insects start feeding on plants with low silica (Massey & Hartley, 2009).

The silicon microstructures slow down insects' penetration into plants, reducing the susceptibility of plants to pathogen damage (Bakhat et al., 2018).

Table 5. Potential mechanisms of the silicon-induced protection against piercing-sucking insect and non-insect pests

Insect Pest	Insect Pest Host plant Mode of action		Reference	
Piercing-sucking pests				
Spider mites	strawberry	Physical barrier, metabolism amendment	Liu et al. (2020)	
Whitefly (Bemisia tabaci)	cucumber	Metabolism amendment	Correa et al. (2005)	
	eggplant	Metabolism amendment, increased growth and Ca intake	Bakhat et al. (2020)	
Aphids Myzus persicae (Sulzer), Sitobion avenae (F.)	wheat	Antibiosis	Dias et al. (2014), Jiang et al. (2023), Ranger et al. (2009)	
Bird cherry-oat aphid <i>Rhopalosiphum padi</i> (Linnaeus)	wheat	Emission of HIPVs	De Oliveira et al. (2020), Liu et al. (2017)	
Corn leaf aphid, <i>Rhopalosiphum maidis</i> (Fitch)	maize	Antibiosis and non-preference	Boer et al. (2019)	

Phytoliths

The physical defense induced by the Si deposition in plant parts in the form of phytoliths (largely composed of SiO₂) was one of the first theories proposed for studying the stress tolerance to pests (Song et al., 2021). The monosilicic acid polymerizes and forms the so called phytoliths, which are accumulated within the plant (Epsteian, 1994). Phytoliths play a structural protective role in plants (Xu et al., 2023). In leaves phytoliths reduce the digestibility of plant tissues and weaken the mouth parts of the pests (Bakhat et al., 2018). The silicon deposition enhances the number of

phytoliths on the surface of stems, which increases the cell wall thickness of the stem sclerenchyma (Miyake & Takahashi 1983). A large number of closely arranged phytoliths protects the vascular tissue of leaves and significantly enhances their resistance (Reynolds et al., 2009).

Papillae

According to Voigt (2014), the papillae are complex structures that are established between the plasma membrane and the inner site of the plant cell wall. The papillae formation is reported as a possible mechanism of the Siinduced defense in plants under biotic stress (Pozza et al., 2015, Verma et al., 2021). These structures could have variable biochemical compositions in the different plant species, but some compounds such as phenolics, reactive oxygen species, cell wall proteins, and cell wall polymers are commonly present.

Callose

Callose is an important linear form of polysaccharide which is synthesized in the plant cell walls. It is mainly build of β -1, 3-linked glucose residues with a small amount of β -1, 6linked branches (Li et al., 2024). Callose deposition is a process that is coordinated through the expression of the genes encoding callose synthase (OsGSL1) and hydrolase (Gns5) (Hao et al., 2008). Callose not only plays an important role in plant development but also participates in the plant defense against environmental stresses (Swaminathan et al., 2022). Other researchers analyze the feeding behavior of phloem-sucking pests and report about the Si induced callose accumulation (Tenguri et al., 2023, Yang et al., 2018). The most intensely silicified tissues are usually the leaf epidermis, root endodermis, and abaxial epidermis of inflorescence bracts (Kumar et al., 2017).

Effect of Si on plant metabolism under biotic stress

The second line of plant defense includes secondary metabolites (such as phenols, lignin) (Alves et al., 2015, Emam et al., 2014, Xue et al., 2022), along with various enzymes (phytohormones) such as polyphenol oxidase (PPO), phenylalanine ammonia lyase (PAL) (Xue et al., 2022) and peroxidase (POD) (Alves et al., 2015), which are involved in their synthesis. One of the mechanisms by which Si is proposed to act and alleviate biotic stress is the production of reactive oxygen species (ROS) and the activation of the antioxidant metabolism (similar to the abiotic stress mitigation) (Van Bockhaven et al., 2013). The generation of ROS and incresed oxidative metabolism help to reduce plants oxidative damage (Domiciano et al, 2015, Yang et al., 2017). These mechanisms have been related to stress due to a pathogen (bacterial and fungal) infection, as well as damage to the plant from piercing-sucking chewing and insects (Domiciano et al, 2015, Yang et al., 2017, Debona et al., 2014). The reactive oxygen species are involved in several signaling actions in different defense signaling pathways with plant hormones, such as JA and SA (Glazebrook, 2005, Ramputh et al., 2002, Torres, 2010, Gulzar et al., 2021). Last but not least, ROS may affect the plant defense genes, resulting in the accumulation of protective metabolic substances such as phytoalexins and allelochemicals in plants (Thoma et al., 2003).

Phenolic and phytoalexin synthesis

The role of phytoalexins in the plant resistance is well established (Dakora & Phillips, 1996, Daniel, 2017, Fawe et al., 1998). Phytoalexins are antimicrobial biomolecules with a low molecular weight that are synthesized in plants as a response to biotic and abiotic stresses (Jeandet, 2015). Silicon can induce the formation of such antimicrobial compounds (Rémus-Borel et al., 2005). Microscopic analyses of the leaf cells of the Sisupplied cucumber plants demonstrated a rapid accumulation of phenolics in response to Podosphaera xanthii infection (Menzies et al., 1991). The high concentrations of flavonoids and phenolic acids in the leaf extracts of the Sitreated cucumber plants reduced powdery mildew symptoms (Emam et al., 2014). The hardness and brittleness of the wounds of Trichothecium roseum-infested muskmelon plants increased during the postharvest period due to lignin, suberin polyphenolic and silicon deposition in the wounded tissue, and the activation of phenylalanine ammonia-lyase (PAL), and the increased content of five phenolic acids (Xue et al., 2022).

Salicylic acid, jasmonic acid, and ethylene synthesis

According to several researchers, Si plays a significant role in the multiple phytohormone signaling pathways to mitigate plant biotic stress (Frew et al., 2018, Manivannan & Ahn, 2017, Coscun et al., 2016). Silicon acts as an emitter of systemic stress signals, leading to the efficient synthesis of protective compounds. The phytohormones salicylic acid (SA), jasmonic acid (JA) and ethylene play a major role in organizing the protective reactions of plants. Jasmonic acid is thought to regulate protection against both chewing insects and piercing-sucking insects, while SA is associated with protection against piercing-sucking insects, feeding on phloem. There is evidence of a strong interaction between Si and JA against insects, which is considered a possible mechanism by which Si increases plant resistance to insect pests.

According to Grant et al. (2013), to protect themselves from microbial pathogen infections, plants have created a multilayer immune system, relaying on constitutive and inducible defense mechanisms, where plant hormones play key roles.

The signaling pathways that allow rice plants to establish resistance to the rice leaf folder (*Cnaphalocrocis medinalis*) are dependent on JA (Liu et al. 2017). Silicon and JA are strongly associated with the different aspects of the rice defense system, as increased levels of transcripts encoding protective genes, the activity of defense-related enzymes (PPO, POD and trypsin protease inhibitor), in addition to the change in the so called herbivore induced plant induced volatiles HIPV (Ye et al., 2013).

Silicon increases the tolerance against the necrotrophic fungal pathogen (*Cochliobolus miyabeanus*), but according to Van Bockhaven et al. (2015) this effect was not dependent on JA and SA pathways. The authors proposed that the Si mode of action is based on the deactivation of the pathogen ethylene production. They suggested that photorespiration and the development of ROS played a vital role in the Si-induced defense against pathogens.

Photosynthesis and enzymatic defense system activation

Photosynthesis is among the most important phenomena, which is responsible for biomass production and overall growth and development in plant (Rastogi et al., 2021). Photosynthesis-related factors play a key role in plant metabolism and are involved in the defense against pathogens (Letousey et al., 2010). There are several studies that investigated the effect of Si in case of pest or disease attacks and proposed that the positive effect of Si was due to the photosynthesis enhancement (Bueno et al., 2017, Sun et al., 2022) and activation of the stress-related enzymes from the antioxidant defense system (Gulzar et al., 2021, Manivannan & Ahn, 2017, Rahman et al. 2015). Silicon pretreatment increased the net photosynthetic rate, and the leaf gas exchange parameters stomatal conductance and transpiration rate in Fusarium oxisporum-infected cucumber (Sun et al., 2022). According to Meunier et al. (2017) one of the reasons for the increased photosynthetic ability of plants is that phytoliths are responsible for the leaf erectness and hence provide large surface sunlight. area to absorb more Si supplementation enhanced the early blight resistance in tomato by modulating the expression of defense-related genes and antioxidant enzymes (Gulzar et al., 2021). Such activation of the antioxidant defense system was observed also in ryegrass against infection by Magnaporthe oryzae in rice (Rahman et al. 2015).

Tritrophic level of silicon-mediated plant defense via HIPVs emission

It is observed that Si is able to influence both bitrophic (plant-herbivore), as well as tritrophic (plant-herbivore–natural enemy) interactions, which, according to Kvedaras et al.

(2010) and Reynolds et al. (2016), may provide another mechanism for pest control. The secondary metabolites are essential in the interactions of plants with insects and other natural enemies. The plant volatile emissions are constitutive or inducible in response to stress and are involved in the protective reactions elicited by grazing animals (Dicke & Baldwin, 2010). In tritrophic systems, the chemical substances are emitted from plants in response to insect induced damage in the form of HIPVs (herbivore induced plant volatiles). These compounds, regardless of their chemical nature, may act either as direct repellents or attractants for the insects and could be used as host signals from entomophagous predators and insect pest parasites (Van Oudenhove, 2017).

Silicon can provoke different plant species to emit, enhance and/ or alter HIPVs (Kvedaras et al. 2010, Alhousari & Greger, 2018). Wild-type rice plants treated with silicon, established a strong indirect protection as a response of the rice leaf folder (Cnaphalocrocis medinalis) feeding, based on HIPVs production (Han et al., 2016, Liu et al., 2017). The volatiles emitted included hexanal 2-ethyl, αbergamoten, β -sesquiophelandrene, and cedrol, produced in significantly smaller amounts in infected Si-treated plants. Their emission significantly enhanced the attraction of adult female parasitoids Trathala flavo-orbitalis and Microplitis mediator to Si-treated plants attacked by C. medinalis (Liu et al. 2017). According to De Oliveira et al. (2020), the silicon-induced changes in the wheat volatile blend mediate the non-preference behavior of the bird cherry-oat aphid (*Rhopalosiphum padi*) and the attraction of the aphid parasitoid Lysiphlebus testaceipes. It was also reported that the Si treatment is able to alter the grapevine volatile profile of the grapevine moth-infested plants (Phalaenoides glycinae Lewin) (Connick, 2011). The authors observed an enhanced production of n-heptadecane, but a decreased synthesis of Cis-thio rose oxide in the Si-supplied plants compared to the controls.

CONCLUSION

Silicon plays an unexpectedly vital role in enhancing the direct and indirect protection of plants against many pests and pathogenes via two closely related mechanisms that, in many cases, act in combination: strengthened physical mechanical barriers and biochemical or mechanisms trigger protective responses in plants. We could speculate that the soil application of silicon may offer a combined physical and chemical protection from diseases and pests, while the foliar application contributes only to the physical defense against enemy penetration. In some cases, the physical amendment ensures more sustainable protection as it is irreversible, and the permanent damages to the pest's digestive system cannot be alleviated via dietary change. Probably, the physical mode of protection is more often observed against chewing pests or fungi. On the other hand, the phloem-feeding piercingsucking pests and bacteria are restricted mostly via the amendment of the plant metabolism. It is hard to judge which protection approach is more effective. Still. reliable and with the development of science and technologies, we could declare that all these mechanisms act synergistically rather than separately, relying on a combination of physical and biochemical amendments to reduce pest and pathogen damage or to attract their natural enemies (Figure 1).

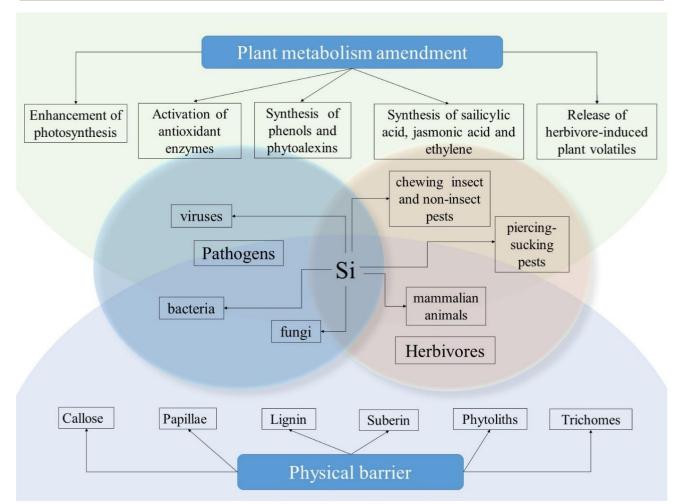


Figure 1. Mechanisms of the silicon-induced defense in plants against pathogenes and herbivores pests

REFERENCES

- Abd-El-Kareem, F., Elshahawy, I. E., & Abd-Elgawad, M. M. (2019). Management of strawberry leaf blight disease caused by Phomopsis obscurans using silicate salts under field conditions. *Bulletin of the National Research Centre*, 43(1), 1-6. <u>https://doi.org/10.1186/s42269-018-</u> 0041-2
- Acevedo, F. E., Peiffer, M., Ray, S., Tan, C. W., & Felton, G. W. (2021). Siliconmediated enhancement of herbivore resistance in agricultural crops. *Frontiers in Plant Science*, 12, 631824.
- Alhousari, F., & Greger, M. (2018). Silicon and mechanisms of plant resistance to insect

pests. *Plants*, 7(2), 33. https://doi.org/10.3390%2Fplants70200 33

- Alves, A. O., Santos, M. M. B., Souza, L. J. N., Souza, E. B., & Mariano, R. L. R. (2015). Use of silicon for reducing the severity of bacterial wilt of sweet pepper. *Journal of Phytopathology*, 97 (3), 419-429.
- Arnon, D. I., & Stout, P. R. (1939). The essentiality of certain elements in minute quantity for plants with special reference to copper. *Plant physiology*, 14(2), 371. <u>https://doi.org/10.1104%2Fpp.14.2.371</u>
- Ashfaq, W., Brodie, G., Fuentes, S., Pang, A., & Gupta, D. (2023). Silicon improves root system and canopy physiology in wheat

under drought stress. *Plant and Soil*, 1-18. <u>https://doi.org/10.1007/s11104-023-</u> 06202-4

- Bakhat, H. F., Bibi, N., Fahad, S., Hammad, H. M., Natasha, Abbas, S., ... & Ashraf, M. R. (2021). Rice husk bio-char improves brinjal growth, decreases insect infestation by enhancing silicon uptake. *Silicon*, 13, 3351-3360. <u>https://doi.org/10.1007/s12633-020-</u>00719-4
- Bakhat, H. F., Bibi, N., Zia, Z., Abbas, S., Hammad, H. M., Fahad, S., ... & Saeed, S. (2018). Silicon mitigates biotic stresses in crop plants: A review. *Crop Protection*, 104, 21-34. <u>https://doi.org/10.1016/j.cropro.2017.10</u> .008
- Ben Rejeb, I., Pastor, V., & Mauch-Mani, B. (2014). Plant responses to simultaneous biotic and abiotic stress: molecular mechanisms. *Plants*, *3*(4), 458-475. <u>https://doi.org/10.3390/plants3040458</u>
- Boer, C. A., Sampaio, M. V., & Pereira, H. S. (2019). Silicon-mediated and constitutive resistance to *Rhopalosiphum maidis* (Hemiptera: Aphididae) in corn hybrids. *Bulletin of entomological research*, 109(3), 356-364.

https://doi.org/10.1017/S000748531800 0585

- Bolton, M. D. (2009). Primary metabolism and plant defense—fuel for the fire. *Molecular plant-microbe Interactions*, 22(5), 487-497. <u>https://doi.org/10.1094/mpmi-22-5-</u> 0487
- Bueno, A. C. S. O., Castro, G. L. S., Silva Junior, D. D., Pinheiro, H. A., Filippi, M. C. C., & Silva, G. B. (2017).
 Response of photosynthesis and chlorophyll a fluorescence in leaf scaldinfected rice under influence of rhizobacteria and silicon fertilizer. *Plant*

pathology, 66(9), 1487-1495. <u>https://doi.org/10.1111/ppa.12690</u>

- Calandra, I., Zub, K., Szafrańska, P. A., Zalewski, A., & Merceron, G. (2016). Silicon-based plant defences, tooth wear and voles. *Journal of Experimental Biology*, 219(4), 501-507.
- Connick, V. J. (2011). The impact of silicon fertilisation on the chemical ecology of grapevine, *Vitis vinifera*; constitutive and induced chemical defences against arthropod pests and their natural enemies. Ph.D. thesis, Charles Sturt University, Albury-Wodonga.
- Correa, R. S., Moraes, J. C., Auad, A. M., & Carvalho, G. A. (2005). Silicon and acibenzolar-S-methyl as resistance inducers in cucumber, against the whitefly *Bemisia tabaci* (Gennadius)(Hemiptera: Aleyrodidae) biotype B. *Neotropical entomology*, 34, 429-433.

http://dx.doi.org/10.1590/S1519-566X2005000300011

- Coskun, D., Britto, D. T., Huynh, W. Q., & Kronzucker, H. J. (2016). The role of silicon in higher plants under salinity and drought stress. *Frontiers in plant science*, 7, 210358. https://doi.org/10.3389/fpls.2016.01072
- Currie, H. A., & Perry, C. C. (2007). Silica in plants: biological, biochemical and chemical studies. *Annals of botany*, 100(7), 1383-1389. <u>https://doi.org/10.1093%2Faob%2Fmc</u> <u>m247</u>
- Dakora, F. D., & Phillips, D. A. (1996). Diverse functions of isoflavonoids in legumes transcend anti-microbial definitions of phytoalexins. *Physiological and Molecular Plant Pathology*, 49(1), 1-20. <u>https://doi.org/10.1006/pmpp.1996.003</u> 5
- Daniel, M. (2017). Handbook of phytoalexin metabolism and action. Routledge.

Agricultural University – Plovdiv 🎇 AGRICULTURAL SCIENCES Volume 16 Issue 41 2024

- Datnoff, L. E., & Rodrigues, F. A. (2015). History of silicon and plant disease. *Silicon and Plant Diseases*, 1-5. <u>https://doi.org/10.1007/978-3-319-</u> 22930-0
- de Oliveira, R. S., Peñaflor, M. F. G., Gonçalves, F. G., Sampaio, M. V., Korndörfer, A. P., Silva, W. D., & Bento, J. M. S. (2020). Silicon-induced changes in plant volatiles reduce attractiveness of wheat to the bird cherry-oat aphid Rhopalosiphum padi and attract the parasitoid *Lysiphlebus testaceipes*. *Plos one*, 15(4), e0231005. <u>https://doi.org/10.1371%2Fjournal.pone</u> .0231005
- de Oliveira, R. S., Sampaio, M. V., Carvalho, F. J., Albuquerque, C. J. B., & Korndorfer, G. H. (2023). Silicon amendments reduce aphid numbers and improve yield of aphid-resistant and susceptible wheat cultivars in a dose-dependent manner. *Crop Protection*, 172, 106296. <u>https://doi.org/10.1016/j.cropro.2023.10</u> 6296
- Debona, D., Rodrigues, F. A., & Datnoff, L. E. (2017). Silicon's role in abiotic and biotic plant stresses. *Annual Review of Phytopathology*, 55, 85-107.
- Debona, D., Rodrigues, F. A., Rios, J. A., Nascimento, K. J. T., & Silva, L. C. (2014). The effect of silicon on antioxidant metabolism of wheat leaves infected by *Pyricularia oryzae*. *Plant pathology*, 63(3), 581-589. <u>https://doi.org/10.1111/ppa.12119</u>
- Dias, P. A. S., Sampaio, M. V., Rodrigues, M. P., Korndörfer, A. P., Oliveira, R. S., Ferreira, S. E., & Korndörfer, G. H. (2014). Induction of resistance by silicon in wheat plants to alate and apterous morphs of Sitobion avenae (Hemiptera: Aphididae). *Environmental entomology*, 43(4), 949-956. https://doi.org/10.1603/EN13234

- Domiciano, G. P., Cacique, I. S., Chagas Freitas, C., Filippi, M. C. C., DaMatta, F. M., do Vale, F. X. R., & Rodrigues, F. Á. (2015). Alterations in gas exchange and oxidative metabolism in rice leaves infected by *Pyricularia oryzae* are attenuated by silicon. *Phytopathology*, 105(6), 738-747. <u>https://doi.org/10.1094/phyto-10-14-</u> 0280-r
- Dos Santos, M. C., Junqueira, M. R., de Sá, V.
 M., Zanúncio, J. C., & Serrão, J. E. (2015). Effect of silicon on the morphology of the midgut and mandible of tomato leafminer *Tuta absoluta* (Lepidoptera: Gelechiidae) larvae. *Invertebrate survival journal*, 12(1), 158-165.
- Emam, M. M., Khattab, H. E., Helal, N. M., & Deraz, A. E. (2014). Effect of selenium and silicon on yield quality of rice plant grown under drought stress. *Australian Journal of Crop Science*, 8(4), 596-605.
- Frew, A., Weston, L. A., Reynolds, O. L., & Gurr, G. M. (2018). The role of silicon in plant biology: a paradigm shift in research approach. *Annals of botany*, 121(7), 1265-1273. https://doi.org/10.1093/aob/mcv009
- Garibaldi, A., Gilardi, G., Cogliati, E. E., & Gullino, M. L. (2012). Silicon and increased electrical conductivity reduce downy mildew of soilless grown lettuce. *European journal of plant pathology*, 132, 123-132. <u>http://dx.doi.org/10.1007%2Fs10658-</u>

<u>011-9855-6</u>

- Glazebrook, J. (2005). Contrasting mechanisms of defense against biotrophic and necrotrophic pathogens. *Annual Review* of *Phytopathology*, 43, 205-227. <u>https://doi.org/10.1146/annurev.phyto.4</u> <u>3.040204.135923</u>
- Gomez, A. O., Mattner, S. W., Oag, D., Nimmo, P., Milinkovic, M., & Villalta, O. N. (2016, August). Protecting fungicide

chemistry used in Australian strawberry production for more sustainable control of powdery mildew and leaf blotch. In *VIII International Strawberry Symposium* 1156, 735-742. <u>https://doi.org/10.17660/ActaHortic.20</u> 17.1156.108

- Grant, M. R., Kazan, K., & Manners, J. M. (2013). Exploiting pathogens' tricks of the trade for engineering of plant disease resistance: challenges and opportunities. *Microbial biotechnology*, 6(3), 212-222. <u>https://doi.org/10.1111/1751-</u> 7915.12017
- Gulzar, N., Ali, S., Shah, M. A., & Kamili, A. N. (2021). Silicon supplementation improves early blight resistance in *Lycopersicon esculentum* Mill. by modulating the expression of defense-related genes and antioxidant enzymes. *3 Biotech*, 11, 1-13. <u>https://doi.org/10.1007/s13205-021-02789-6</u>
- Guntzer, F., Keller, C., & Meunier, J. D. (2012). Benefits of plant silicon for crops: a review. Agronomy for sustainable development, 32, 201-213. <u>https://doi.org/10.1007/s13593-011-</u>0039-8
- Han, Y., Li, P., Gong, S., Yang, L., Wen, L., & Hou, M. (2016). Defense responses in rice induced by silicon amendment against infestation by the leaf folder Cnaphalocrocis medinalis. *PloS one*, 11(4), e0153918. <u>https://doi.org/10.1371/journal.pone.01</u> 53918
- Hao, P., Liu, C., Wang, Y., Chen, R., Tang, M., Du, B., ... & He, G. (2008). Herbivoreinduced callose deposition on the sieve plates of rice: an important mechanism for host resistance. *Plant physiology*, 146(4), 1810-1820. https://doi.org/10.1104/pp.107.111484
- Haq, I. U., Zhang, K., Ali, S., Majid, M., Ashraf, H. J., Khurshid, A., ... & Liu, C. (2022).

Effectiveness of silicon on immature stages of the fall armyworm [*Spodoptera frugiperda* (JE Smith)]. *Journal of King Saud University-Science*, 34(6), 102152. https://doi.org/10.1016/j.jksus.2022.102 152

- Hernandez-Apaolaza, L. (2014). Can silicon partially alleviate micronutrient deficiency in plants? A review. *Planta*, 240(3), 447-458. <u>https://doi.org/10.1007/s00425-014-</u> 2119-x
- Hodson, M. J. (1986). Silicon deposition in the roots, culm and leaf of *Phalaris canariensis* L. *Annals of Botany*, 58(2), 167-177.
 https://doi.org/10.1093/oxfordjournals.a ob.a087194
- https://doi.org/10.1007/s00425-003-1105-5
- IPNI, 2015. Silicon. Nutri-facts, No. 14. <u>http://www.ipni.net/publication/nutrifac</u> <u>ts-</u> <u>na.nsf/0/A7B4AB4D35C153BF85257E</u> <u>CE006E0E34/\$FILE/NutriFacts-NA-</u> 14.pdf
- Jeandet, P. (2015). Phytoalexins: current progress and future prospects. *Molecules*, 20(2), 2770-2774. <u>https://doi.org/10.3390%2Fmolecules20</u> 022770
- Jiang, Y., Qi, X. X., Wang, Z. H., Liu, X. D., Han, Y. L., Li, H., & Wang, Y. (2023). Effect of Different Forms of Silicon Application on Wheat Aphid Resistance. *Agricultural Research*, 12(2), 179-188.
- Jinger, D., Devi, M. T., Dhar, S., Dass, A., Rajanna, G. A., Upadhaya, P., & Raj, R. (2017). Silicon in mitigating biotic stresses in rice (*Oryza sativa* L.)" a review. Annals of Agricultural
- Kanto, T., Maekawa, K., & Aino, M. (2007). Suppression of conidial germination and appressorial formation by silicate treatment in powdery mildew of strawberry. *Journal of General Plant Pathology*, 73(1), 1-7.

https://doi.org/10.1007/s10327-006-0311-y

- Kumar, R., & Rathor, V. S. (2020). Nature and types of damage by insect pests. *Journal of Entomological Research*, 44(4), 639-646.
- Kumar, S., Soukup, M., & Elbaum, R. (2017). Silicification in grasses: variation between different cell types. *Frontiers in Plant Science*, 8, 259110. https://doi.org/10.3389/fpls.2017.00438
- Kvedaras, O. L., An, M., Choi, Y. S., & Gurr, G. M. (2010). Silicon enhances natural enemy attraction and biological control through induced plant defences. *Bulletin* of entomological research, 100(3), 367-371.

https://doi.org/10.1017/s000748530999 0265

- Letousey, P., Baillieul, F., Perrot, G., Rabenoelina, F., Boulay, M., Vaillant-Gaveau, N., ... & Fontaine, F. (2010). Early events prior to visual symptoms in the apoplectic form of grapevine esca disease. *Phytopathology*, 100(5), 424-431. <u>https://doi.org/10.1094/phyto-100-5-0424</u>
- Li, N., Lin, Z., Yu, P., Zeng, Y., Du, S., & Huang, L. J. (2023). The multifarious role of callose and callose synthase in plant development and environment interactions. *Frontiers in Plant Science*, 14, 1183402. <u>https://doi.org/10.3389/fpls.2023.11834</u> 02
- Li, Y. C., Bi, Y., Ge, Y. H., Sun, X. J., & Wang, Y. (2009). Antifungal activity of sodium silicate on Fusarium sulphureum and its effect on dry rot of potato tubers. *Journal of food Science*, 74(5), 213-218. <u>https://doi.org/10.1111/j.1750-3841.2009.01154.x</u>
- Liu, B., Davies, K., & Hall, A. (2020). Silicon builds resilience in strawberry plants against both strawberry powdery mildew *Podosphaera aphanis* and two-

spotted spider mites *Tetranychus urticae*. *PLoS One*, 15(12), e0241151. <u>https://doi.org/10.1371/journal.pone.02</u> <u>41151</u>

- Liu, J., Zhu, J., Zhang, P., Han, L., Reynolds, O.
 L., Zeng, R., ... & Gurr, G. M. (2017).
 Silicon supplementation alters the composition of herbivore induced plant volatiles and enhances attraction of parasitoids to infested rice plants. *Frontiers in plant science*, 8, 1265.
 <u>https://doi.org/10.3389/fpls.2017.01265</u>
- Ma, J. F., & Yamaji, N. (2015). A cooperative system of silicon transport in plants. *Trends in plant science*, 20(7), 435-442. <u>https://doi.org/10.1016/j.tplants.2015.0</u> <u>4.007</u>
- Manivannan, A., & Ahn, Y. K. (2017). Silicon regulates potential genes involved in major physiological processes in plants to combat stress. *Frontiers in Plant Science*, 8, 275701. https://doi.org/10.3389/fpls.2017.01346
- Massey, F. P., & Hartley, S. E. (2009). Physical defences wear you down: progressive and irreversible impacts of silica on insect herbivores. *Journal of animal ecology*, 78(1), 281-291. <u>https://doi.org/10.1111/j.1365-</u> 2656.2008.01472.x
- Massey, F. P., Ennos, A. R., & Hartley, S. E. (2006). Silica in grasses as a defence against insect herbivores: contrasting effects on folivores and a phloem feeder. *Journal of Animal Ecology*, 75(2), 595-603. <u>https://doi.org/10.1242/jeb.134890</u>
- Menzies, J. G., Ehret, D. L., Glass, A. D. M., & Samuels, A. L. (1991). The influence of silicon on cytological interactions between Sphaerotheca fuliginea and *Cucumis sativus*. *Physiological and Molecular Plant Pathology*, 39(6), 403-414. <u>https://doi.org/10.1016/0885-5765(91)90007-5</u>
- Meunier, J. D., Barboni, D., Anwar-ul-Haq, M., Levard, C., Chaurand, P., Vidal, V., ... &

Agricultural University – Plovdiv 🎇 AGRICULTURAL SCIENCES Volume 16 Issue 41 2024

Keller, C. (2017). Effect of phytoliths for mitigating water stress in durum wheat. New Phytologist, 215(1), 229-239. <u>https://doi.org/10.1111/nph.14554</u>

- Miyake, Y., & Takahashi, E. (1983). Effect of silicon on the growth of solutioncultured cucumber plant. *Soil Science and Plant Nutrition*, 29(1), 71-83. <u>https://doi.org/10.1080/00380768.1983.</u> 10432407
- Mordecai, E. A. (2011). Pathogen impacts on plant communities: unifying theory, concepts, and empirical work. *Ecological Monographs*, 81(3), 429-441. <u>https://doi.org/10.1890/10-2241.1</u>
- Nascimento, A. M., Assis, F. A., Moraes, J. C., & Souza, B. H. S. (2018). Silicon application promotes rice growth and negatively affects development of Spodoptera frugiperda (JE Smith). *Journal of applied entomology*, 142(1-2), 241-249.
- Otitodun, G. O., Opit, G. P., Nwaubani, S. I., & Okonkwo, E. U. (2017). Efficacy of rice husk ash against rice weevil and lesser grain borer on stored wheat. African Crop Science Journal,
- Parrilli, M., Sommaggio, D., Tassini, C., Di Marco, S., Osti, F., Ferrari, R., ... & Burgio, G. (2019). The role of Trichoderma spp. and silica gel in plant defence mechanisms and insect response in vineyard. *Bulletin of entomological research*, 109(6), 771-780. <u>https://doi.org/10.1017/s000748531900</u> 0075
- Pozza, E. A., Pozza, A. A. A., & Botelho, D. M. D. S. (2015). Silicon in plant disease control. *Revista Ceres*, 62, 323-331. <u>https://doi.org/10.1590/0034-</u> 737X201562030013
- Rahman, A., Wallis, C. M., & Uddin, W. (2015). Silicon-induced systemic defense responses in perennial ryegrass against infection by *Magnaporthe oryzae*. *Phytopathology*, 105(6), 748-

757. <u>https://doi.org/10.1094/PHYTO-</u> 12-14-0378-R

- Ramputh, A. I., Arnason, J. T., Cass, L., & Simmonds, J. A. (2002). Reduced herbivory of the European corn borer (*Ostrinia nubilalis*) on corn transformed with germin, a wheat oxalate oxidase gene. *Plant Science*, 162(3), 431-440. <u>https://doi.org/10.1016/S0168-</u> 9452(01)00584-2
- Rastogi, A., Yadav, S., Hussain, S., Kataria, S., Hajihashemi, S., Kumari, P., ... & Brestic, M. (2021). Does silicon really matter for the photosynthetic machinery in plants...? *Plant Physiology and Biochemistry*, 169, 40-48. https://doi.org/10.1016/j.plaphy.2021.1 1.004
- Rémus-Borel, W., Menzies, J. G., & Bélanger, R. R. (2005). Silicon induces antifungal compounds in powdery mildew-infected wheat. *Physiological and molecular plant pathology*, 66(3), 108-115.
- Resende, R. S., Rodrigues, F. Á., Costa, R. V., & Silva, D. D. (2013). Silicon and fungicide effects on anthracnose in moderately resistant and susceptible sorghum lines. *Journal of Phytopathology*, 161(1), 11-17. <u>https://doi.org/10.1111/jph.12020</u>
- Reynolds, O. L., Keeping, M. G., & Meyer, J. H. (2009). Silicon-augmented resistance of plants to herbivorous insects: a review. *Annals of applied biology*, 155(2), 171-186. <u>http://dx.doi.org/10.1111/j.1744-</u> 7348.2009.00348.x
- Reynolds, O. L., Padula, M. P., Zeng, R., & Gurr, G. M. (2016). Silicon: potential to promote direct and indirect effects on plant defense against arthropod pests in agriculture. *Frontiers in plant science*, 7, 188037.

https://doi.org/10.3389/fpls.2016.00744

Richmond, K. E., & Sussman, M. (2003). Got silicon? The non-essential beneficial

plant nutrient. *Current opinion in plant biology*, 6(3), 268-272. https://doi.org/10.1016/S1369-5266(03)00041-4

- Rodrigues, F. A., Duarte, H. S. S., Domiciano,
 G. P., Souza, C. A., Korndörfer, G. H.,
 & Zambolim, L. (2009). Foliar application of potassium silicate reduces the intensity of soybean rust. *Australasian Plant Pathology*, 38(4), 366-372.
- Roy, S., Mohammad, R., Khamari, B., Monalisa, S. P., & Swain, D. K. (2023).
 Silicon mediated defense response in rice plants against brown plant hopper *Nilaparvata lugens* (Stål). *Silicon*, 15(17), 7579-7591.
 <u>https://doi.org/10.1007/s12633-023-</u> 02610-4
- Sakr, N. (2021). Silicon reduces Fusarium head blight development in barley. The Open *Agriculture Journal*, 15(1), 54-65. <u>http://dx.doi.org/10.2174/18743315021</u> <u>15010054</u>
- Samuels, A. L., Glass, A. D. M., Menzies, J. G., & Ehret, D. L. (1994). Silicon in cell walls and papillae of *Cucumis sativus* during infection by *Sphaerotheca fuliginea*. *Physiological and molecular plant pathology*, 44(4), 237-242. <u>https://doi.org/10.1016/S0885-</u> <u>5765(05)80027-X</u>
- Savvas, D., Giotis, D., Chatzieustratiou, E., Bakea, M., & Patakioutas, G. (2009). Silicon supply in soilless cultivations of zucchini alleviates stress induced by salinity and powdery mildew infections. *Environmental and experimental botany*, 65(1), 11-17. <u>https://doi.org/10.1016/j.envexpbot.200</u> 8.07.004
- Schuerger, A. C., & Hammer, W. (2003). Suppression of powdery mildew on greenhouse-grown cucumber by addition of silicon to hydroponic nutrient solution is inhibited at high

temperature. *Plant Disease*, 87(2), 177-185.

https://doi.org/10.1094/pdis.2003.87.2.1 77

- Seal, P., Das, P., & Biswas, A. K. (2018). Versatile potentiality of silicon in mitigation of biotic and abiotic stresses in plants: a review. *American Journal of Plant Sciences*, 9(7), 1433-1454. https://doi.org/10.4236/ajps.2018.97105
- Shwethakumari, U., & Prakash, N. B. (2018). Effect of foliar application of silicic acid on soybean yield and seed quality under field conditions. *Journal of the Indian Society of Soil Science*, 66(4), 406-414.
- Sidhu, J. K., Stout, M. J., Blouin, D. C., & Datnoff, L. E. (2013). Effect of silicon soil amendment on performance of sugarcane borer, *Diatraea saccharalis* (Lepidoptera: Crambidae) on rice. *Bulletin of entomological research*, 103(6), 656-664. <u>https://doi.org/10.1017/s000748531300</u> 0369
- Singh, P., Kumar, V., Sharma, J., Saini, S., Sharma, P., Kumar, S., ... & Sharma, A. Silicon supplementation (2022).alleviates the salinity stress in wheat plants by enhancing the plant water status, photosynthetic pigments, proline content and antioxidant enzyme 2525. activities. Plants, 11(19), https://doi.org/10.3390/plants11192525
- Somapala, K., Weerahewa, D., & Thrikawala, S. (2016). Silicon rich rice hull amended soil enhances anthracnose resistance in tomato. *Procedia food science*, 6, 190-193.

https://doi.org/10.1016/j.profoo.2016.0 2.046

Song, A., Xue, G., Cui, P., Fan, F., Liu, H., Yin, C., ... & Liang, Y. (2016). The role of silicon in enhancing resistance to bacterial blight of hydroponic-and soilcultured rice. Scientific Reports, 6(1), 24640.

https://doi.org/10.1038/srep24640

- Song, X. P., Verma, K. K., Tian, D. D., Zhang, X. Q., Liang, Y. J., Huang, X., ... & Li, Y. R. (2021). Exploration of silicon functions to integrate with biotic stress tolerance and crop improvement. *Biological Research*, 54. <u>https://doi.org/10.1186/s40659-021-</u>00344-4
- Sun, S., Yang, Z., Song, Z., Wang, N., Guo, N., Niu, J., ... & Chen, S. (2022). Silicon enhances plant resistance to Fusarium wilt by promoting antioxidant potential and photosynthetic capacity in cucumber (*Cucumis sativus* L.). *Frontiers in Plant Science*, 13, 1011859. <u>https://doi.org/10.3389/fpls.2022.10118</u> 59
- Swaminathan, S., Lionetti, V., & Zabotina, O. A. (2022). Plant cell wall integrity perturbations and priming for defense. *Plants*, 11(24), 3539. <u>https://doi.org/10.3390/plants11243539</u>
- Tenguri, P., Chander, S., Ellur, R. K., Yele, Y., Sundaran, A. P., Nagaraju, M. T., ... & Suroshe, S. S. (2023). Effect of silicon application to the rice plants on feeding behaviour of the brown planthopper, *Nilaparvata lugens* (Stål) under elevated CO₂. Silicon, 15(13), 5811-5820. <u>https://doi.org/10.1007/s12633-023-</u> 02480-w
- Thoma, I., Loeffler, C., Sinha, A. K., Gupta, M., Krischke, M., Steffan, B., ... & Mueller, M. J. (2003). Cyclopentenone isoprostanes induced by reactive oxygen species trigger defense gene activation and phytoalexin accumulation in plants. *The Plant Journal*, 34(3), 363-375. <u>https://doi.org/10.1046/j.1365-</u> 313x.2003.01730.x
- Torres, M. A. (2010). ROS in biotic interactions. *Physiologia plantarum*, 138(4), 414-429.

https://doi.org/10.1111/j.1399-3054.2009.01326.x

- Van Bockhaven, J., De Vleesschauwer, D., & Höfte, M. (2013). Towards establishing broad-spectrum disease resistance in plants: silicon leads the way. *Journal of experimental botany*, 64(5), 1281-1293. <u>https://doi.org/10.1093/jxb/ers329</u>
- Van Bockhaven, J., Spíchal, L., Novák, O., Strnad, M., Asano, T., Kikuchi, S., ... & De Vleesschauwer, D. (2015). Silicon induces resistance to the brown spot fungus *Cochliobolus miyabeanus* by preventing the pathogen from hijacking the rice ethylene pathway. *New Phytologist*, 206(2), 761-773. <u>https://doi.org/10.1111/nph.13270</u>
- Verma, K. K., Song, X. P., Tian, D. D., Guo, D. J., Chen, Z. L., Zhong, C. S., ... & Li, Y. R. (2021). Influence of silicon on biocontrol strategies to manage biotic stress for crop protection, performance, and improvement. *Plants*, 10(10), 2163. <u>https://doi.org/10.3390/plants10102163</u>
- Vicari, M., & Bazely, D. R. (1993). Do grasses fight back? The case for antiherbivore defences. *Trends in ecology & evolution*, 8(4), 137-141. <u>https://doi.org/10.1016/0169-</u> 5347(93)90026-1
- Vilela, M., Moraes, J. C., Alves, E., Santos-Cividanes, T. M., & Santos, F. A. (2014). Induced resistance to *Diatraea* saccharalis (Lepidoptera: Crambidae) via silicon application in sugarcane. *Revista Colombiana de Entomología*, 40(1), 44-48.
- Voigt, C. A. (2014). Voigt, C. A. (2014). Callose-mediated resistance to pathogenic intruders in plant defenserelated papillae. *Frontiers in plant science*, 5, 86993. https://doi.org/10.3389/fpls.2014.00168
- Wang, W., Vinocur, B., & Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic

engineering for stress tolerance. *Planta*, 218, 1-14.

- White, W. H., & White, P. J. (2013). Sugarcane borer resistance in sugarcane as affected by silicon applications in potting medium, *Journal of the American Society of Sugar Cane*. 38-54.
- Xu, R., Huang, J., Guo, H., Wang, C., & Zhan, H. (2023). Functions of silicon and phytolith in higher plants. *Plant Signaling & Behavior*, 18(1), 2198848. <u>https://doi.org/10.1080/15592324.2023.</u> 2198848
- Xue, G., Sun, W., Song, A., Li, Z., Fan, F., & Liang, Y. (2010). Influence of silicon on rice growth, resistance to bacterial blight and activity of pathogenesis-related proteins. *Scientia Agricultura Sinica*, 43(4), 690-697.
- Xue, S., Li, Z., Li, B., Ackah, S., Wang, B., Zheng, X., ... & Prusky, D. (2022). Sodium silicate treatment promotes suberin poly phenolic and silicon deposition, and enhances hardness and brittleness at muskmelon wounds. *Scientia Horticulturae*, 300, 111087. <u>https://doi.org/10.1016/j.scienta.2022.1</u> 11087
- Yang, L., Han, Y., Li, P., Li, F., Ali, S., & Hou, M. (2017). Silicon amendment is involved in the induction of plant defense responses to a phloem feeder. *Scientific Reports*, 7(1), 4232. <u>https://doi.org/10.1038/s41598-017-04571-2</u>
- Yang, L., Li, P., Li, F., Ali, S., Sun, X., & Hou, M. (2018). Silicon amendment to rice plants contributes to reduced feeding in a phloem-sucking insect through modulation of callose deposition. *Ecology and Evolution*, 8(1), 631-637. <u>https://doi.org/10.1002/ece3.3653</u>
- Yoshida, S., Ohnishi, Y., & Kitagishi, K. (1962). Histochemistry of silicon in rice plant: III. The presence of cuticle-silica double layer in the epidermal tissue. *Soil*

Science and Plant Nutrition, 8(2), 1-5. <u>https://doi.org/10.1080/00380768.1962.</u> 10430982

Zellner, W., Frantz, J., & Leisner, S. (2011). Silicon delays Tobacco ringspot virus systemic symptoms in *Nicotiana tabacum. Journal of Plant Physiology*, 168(15), 1866-1869. <u>https://doi.org/10.1016/j.jplph.2011.04.</u> 002